

The HPLC method for fast determination of levofloxacin in liquid pharmaceutical formulations

Metoda HPLC do szybkiego oznaczenia lewofloksacyny w płynnych postaciach leku

Andrzej Czyrski

The Department of Physical Pharmacy and Pharmacokinetics Poznań University of Medical Sciences

Abstract

Background. Levofloxacin is a representative of the third generation of fluoroquinolones. It is an antimicrobial agent with a wide spectrum of the bactericidal activity. It can be found in liquid pharmaceutical formulations such as infusions or eye drops. This paper reports a fast HPLC method for determination of levofloxacin in liquid pharmaceutical formulations. **Material and methods.** The applied method is RP-HPLC method with UV detection. The sample pre-conditioning involves simple dilution. The calibration curve range obeys 1-6 mg/mL and LOD is 0.019 mg/mL. **Results.** The validation parameters did not exceed 5% and 2% for interday and intraday assay respectively. The analyte remained stable within the analysis and after the long term of storage. The time of the analysis is very fast – 5 min. **Conclusions.** The reported method obeys the validation parameters and is suitable for drug analysis. It provides fast and reliable results. (*Farm Współ 2018; 11: 67-71*)

Keywords: levofloxacin, HPLC, infusions, eye-drops, drug analysis

Streszczenie

Wstęp. Lewofloksacyna jest reprezentatem III generacji fluorochinolonów. Posiada szerokie spektrum działania przeciwbakteryjnego. Na rynku dostępna jest w postaci tabletek oraz płynnych postaci leku takich jak wlewy dożylny oraz krople do oczu. Niniejszy artykuł opisuje szybką metodę do oznaczania lewofloksacyny w płynnych postaciach leku. **Materiał i metody.** Zastosowana została metoda HPLC w odwróconym układzie faz z detekcją UV. Przygotowanie próbek do analizy obejmowało rozcieńczenie. Zakres krzywej wzorcowej obejmował 1-6 mg/mL a LOD wynosił 0,019 mg/mL. **Wyniki.** Parametry walidacyjne nie przekraczały 5% oraz 2% dla oznaczeń *interday* oraz *intraday*. Analit pozostał stabilny w trakcie trwania analizy oraz podczas długiego czasu przechowywania. Czas analizy był bardzo krótki – 5 minut. **Wnioski.** Opracowana metoda spełnia wymagania walidacyjne i jest właściwa do analizy leku. Zapewnia szybkie i wiarygodne wyniki. (*Farm Współ 2018; 11: 67-71*)

Słowa kluczowe: lewofloksacyna, HPLC, wlew dożylny, krople do oczu, analiza leku

Introduction

Levofloxacin (LEVO) is an antimicrobial agent which is a representative of the third generation of the fluoroquinolones. It is an L-isomer of ofloxacin. It has a broad bactericidal activity against many Gram-positive, Gram-negative and also atypical bacteria such as Legionella and Mycoplasma. The activity of LEVO is based on the inhibition of the enzymes that are responsible for separation of bacterial DNA i.e. topoisomerase IV and gyrase. It results in the inhibition of cell replication and, in consequence, the cell death [1-4].

The effectiveness of activity of LEVO, as well as the other representatives of fluoroquinolones, strongly depends on its concentration in blood. The bactericidal effect is provided with a repeatable dose in a defined time interval. It prevents the resistance of the bacteria to the treatment. The other factor that may influence the effectiveness of the treatment is the stability of the substance in a pharmaceutical formulation. LEVO is available on the market in liquid pharmaceutical formulations such as infusions and eye-drops. This paper describes the rapid, simple, selective and precise

HPLC method for determination of LEVO in liquid formulations.

Material and methods

HPLC conditions

Chromatographic analysis was performed at the room temperature. The LiChroCART column (125 × 4 mm, 5 µm, Merck, Germany) with LiChroCART guard column (4 × 4 mm, 5 µm, Merck Germany) were applied for chromatographic separation. The separation was performed in a chromatograph 1220 Infinity LC (Agilent Technologies). The mobile phase consisted of acetonitrile (HPLC grade, Merck, Germany) and 0.4% aqueous solution of triethylamine (POCH, Gliwice, Poland) (pH 3.0) solutions in the following ratio (24:76 v/v) and it was degassed by ultrasonic degassing. The pH of triethylamine solution was achieved by an addition of concentrated phosphorous acid (Fluka, Germany). The flow rate was 1 mL/min, and the detection wavelength was $\lambda=295$ nm. The data were analysed with OpenLab ChemStation ver. A.01.05 software.

Calibration standards

The following stock solutions were prepared: LEVO (Sigma-Aldrich, Germany) and moxifloxacin (IS) (Santa Cruz Biotechnology, USA). The concentration of LEVO was 12 mg/mL and for IS was 1 mg/mL. The substances were dissolved in methanol. The standard solutions of LEVO with the following concentrations were prepared: 1 mg/mL, 2 mg/mL, 3 mg/mL, 4 mg/mL, 5 mg/mL and 6 mg/mL. The IS solution concentration was 150 mg/L.

Sample preparation

The matrices of a liquid pharmaceutical formulations consisted mainly of isotonic solution of sodium chloride. The concentration of LEVO in the formula-

tions was 5 mg/mL. 10 µL of the investigated infusion was added to 990 µL of methanol. 100 µL of the dilution was mixed with 20 µL of IS solution and filled with methanol to 200 µL. The 20 µL of the final mixture was injected onto HPLC system.

The validation parameters

The following validation parameters were assayed: linearity, precision and accuracy.

The linear equation describing the relationship between LEVO concentration and the area under the peak of LEVO and IS ratio. The calibration curve was calculated by the least squares method. Five calibration curves were prepared on five separate days and the validation parameters were calculated.

For the inter- and intra-day assay precision and accuracy for low (1 mg/mL, 2 mg/mL) and high concentrations (5 mg/mL, 6 mg/mL) quality samples were analysed (Table I).

The stability was evaluated during three freeze-thaw cycles and after storage at a room temperature 24 h. The analytes are stable if the deviation from the nominal concentration is within $\pm 10\%$.

Limit of detection (LOD)

The limit of detection was calculated from the following equation:

$$LOD = \frac{3.3 \times s}{b}$$

s-standard deviation of the interception, b- slope of the equation curve

Results and discussion

The fast method for the determination of the drug is useful in drug analysis. The mobile phase consisted of acetonitrile and triethylamine (24:76, v/v) solution which was an ion pair reagent. This proportion provides

Table I. The validation parameters

Tabela I. Parametry walidacji

| Concentration [mg/mL] | Inter-day | | Intra-day | |
|-----------------------|-----------|----------|-----------|----------|
| | Precision | Accuracy | Precision | Accuracy |
| 1 | 4.21 | 2.15 | 1.74 | 0.21 |
| 2 | 3.17 | 3.01 | 1.61 | 0.86 |
| 5 | 2.11 | 1.75 | 1.12 | 1.61 |
| 6 | 3.84 | 0.91 | 0.15 | 1.17 |

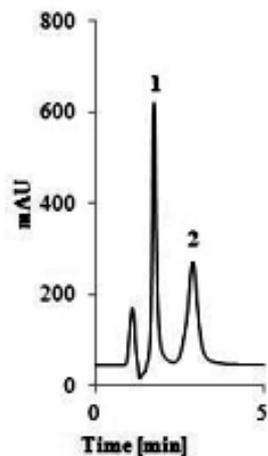


Figure 1. The chromatogram of the sample of commercially available infusion. The concentration of levofloxacin 5 mg/mL (levofloxacin (1) tr- 2 min and IS (2) tr – 3.7 min)

Rycina 1. Chromatogram próby pobranej z komercyjnie dostępnego preparatu do wlewu dożylnego. Stężenie lewofloksacyny 5 mg/mL (lewofloksacyna (1) tr – 2 min and IS (2) tr – 3,7 min)

the total resolution of the LEVO and IS. The chromatographic profile is shown in Fig. 1. The retention time for levofloxacin is 2 min. and IS is 3.7 min. This method characterizes a short time of analysis – 5 min.

The addition of ion pair reagent causes a better interaction of LEVO with the stationary phase and it reduces the tailing of the peaks when combined with the proper value of pH. The separation mechanism is an interaction of LEVO and with the free silanol groups of the stationary phase. Triethylamine reduces the availability of these groups for LEVO and it resulted in reducing of the tailing effect. The silanol groups are ionized at the pH above 3.5 and they interact with 1° and 2° amines. The pH of the mobile phase was 3.0. The increase of pH resulted in the peak tailing. According to the literature data, the concentration of ion pair reagent up to 1% combined with a slightly acidic pH provides a good resolution of the analytes [5,6]. In our study, 0.4% triethylamine combined with a pH value 3.0 resulted in a total resolution of the analytes. Too high concentration of the ion pair reagent results in a long time of the column equilibration. The other factor that was considered for improving the shape of the peak was the phosphate buffer, however it did not improve the quality of the resolution of analytes. The applied organic solvent was acetonitrile. The use of methanol resulted in longer time of analysis. The peaks were

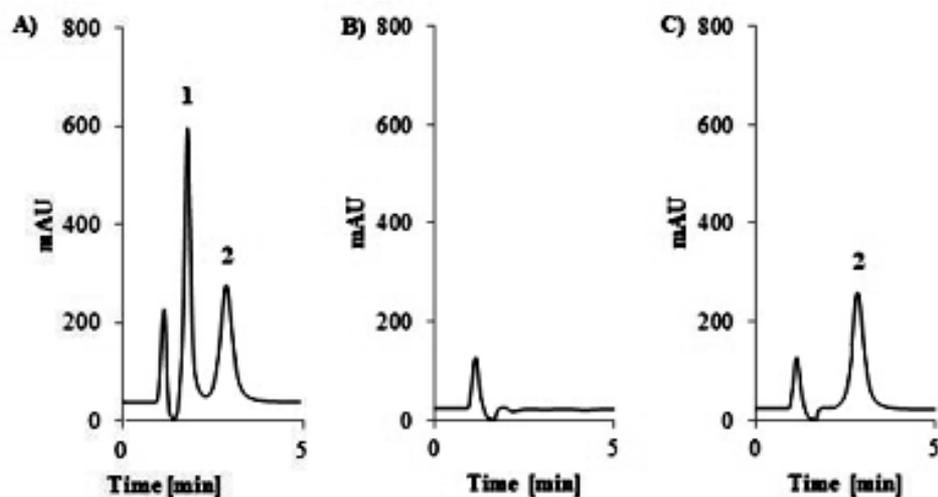


Figure 2. The chromatogram of the: A) calibration curve sample (LEVO concentration 5mg/mL), B) blind sample, C) zero sample, where 1- levofloxacin, 2- IS

Rycina 2. Chromatogram: A) próbki z krzywej wzorcowej (stężenie LEVO 5mg/mL), B) próby ślepej, C) próby zerowej, gdzie 1-lewofloksacyna, 2- IS

broad and the pressure on the column was high, the resolution was poor. The mixtures of acetonitrile and methanol in different proportions were also tested but it resulted in poor resolution. The chromatogram of the sample of the calibration curve, the zero and blind sample was showed in Fig 2.

Many separation methods were reported for LEVO determination in pharmaceutical formulation. They involved capillary electrophoresis [7], HPLC with ultraviolet detection [8, 9]. The developed method was a modification of the method developed by the Santoro et al. [9], however in our study the internal standard was used. The method was tested on the commercially available infusion of levofloxacin at the concentration 5 mg/mL and the separation provided the total resolution of the analyte and IS (Fig.1). The temperature was ambient. The higher temperature does not influence the chromatographic separation significantly [10].

The validation parameters obey the ICH recommendations for bioanalytical methods. The inter-day precision and accuracy do not exceed the value of 4.21% and 3.01 respectively. The intra-day precision and accuracy do not exceed the value 1.74% and 1.61% respectively (Table I).

The sample pre-conditioning involved a simple dilution with methanol. The matrix was not complex and it was not necessary to apply more laborious techniques of sample preparation such as liquid-liquid extraction or solid-phase extraction. The dilution of the liquid formulation was also applied by Gupta et al. [11], Kalarya et al. [5] and Razzaq et al. [12].

The analyte remained stable in stability tests. After storage for 24 hours at the room temperature and after storage at the temperature -20°C for three months the

accuracy (denoted as RE) did not exceed 2.0%. After three freeze-thaw-cycles the accuracy did not exceed 2.1% (Table II).

The calibration curve was linear within the range 1-6 mg/mL. The regression coefficient was higher than 0.999. The test value in Mandell's test was 6.35 vs. the critical value 34.11 and it also confirmed the linearity.

The lowest concentration from the calibration curve is 1 mg/mL. The LOD is 0.019 mg/mL and it is satisfactory for the determination of LEVO in infusions.

Conclusions

In this study, the fast and cheap method for determination of LEVO was developed. The method fulfills the validation parameters for the analytical methods and can be applied in drug analysis.

Funding

This study was supported by the Poznan University of Medical Sciences. Grant number: 502-14-03306416-08887

Conflict of interest

None

Correspondence address

✉ Andrzej Czyrski
The Department of Physical Pharmacy
and Pharmacokinetics
Poznań University of Medical Sciences,
6, Święcickiego St.; 60-781 Poznań, Poland
☎ (+48 61) 854 64 33
✉ aczyrski@ump.edu.pl

Table II. The stability of levofloxacin

Tabela II. Stabilność lewofloksacyny

| Experimental conditions | Concentration, mg/mL | | | |
|---|----------------------|------|------|------|
| | 1.0 | 2.0 | 5.0 | 6.0 |
| <i>After storage at room temperature for 24 h</i> | | | | |
| Mean assayed value, mg/mL | 0.97 | 2.02 | 5.11 | 6.11 |
| Accuracy, %RE | 2.0 | 0.5 | 2.0 | 0.2 |
| <i>After three freeze-thaw cycles</i> | | | | |
| Mean assayed value, mg/mL | 1.02 | 2.03 | 5.04 | 6.04 |
| Accuracy, %RE | 1.0 | 2.1 | 1.4 | 0.8 |
| <i>After storage at -20°C for three months</i> | | | | |
| Mean assayed value, mg/mL | 1.03 | 2.04 | 5.08 | 6.06 |
| Accuracy, %RE | 2.0 | 1.5 | 1.6 | 0.7 |

References

- 1 Hawkey PM. Mechanisms of quinolone action and microbial response. *J Antimicrob Chemother.* 2003;51(Suppl 1):29-35.
- 2 Ferrara AM. New fluoroquinolones in lower respiratory tract infections and emerging patterns of pneumococcal resistance. *Infection.* 2005;33:106-14.
- 3 Appelbaum PC, Hunter PA. The fluoroquinolone antibacterials: past, present and future perspectives. *Int J Antimicrob Ag.* 2000;16:5-15.
- 4 Czyrski A, Kondys K, Szalek E, et al. The pharmacokinetic interaction between levofloxacin and sunitinib. *Pharmacol Rep.* 2015;67(3):542-44.
- 5 Kalariya PD, Namdev D, Srinivas R, et al. Application of Experimental Design and Response Surface Technique for Selecting the Optimum RP-HPLC Conditions for the Determination of Moxifloxacin HCl and Ketorolac Tromethamine in Eye Drops. *J Saudi Chem Soc.* 2017;21(1):S373-S382.
- 6 Czyrski A. Analytical Methods for Determining Third and Fourth Generation Fluoroquinolones: A Review. *Chromatographia.* 2017;80(2):181-200.
- 7 Faria AF, de Souza MVN, de Almeida MV, et al. Simultaneous separation of five fluoroquinolone antibiotics by capillary zone electrophoresis. *Anal Chim Acta.* 2006;579(2):185-92.
- 8 Dafale NA, Semwal UP, Agarwal PK, et al. Development and validation of microbial bioassay for quantification of levofloxacin in pharmaceutical preparations. *J Pharm Anal.* 2015;5:18-26.
- 9 Santoro M, Kassab N, Singh A, et al. Quantitative determination of gatifloxacin, levofloxacin, lomefloxacin and pefloxacin fluoroquinolonic antibiotics in pharmaceutical preparations by high-performance liquid chromatography. *J Pharm Biomed Anal.* 2006;40:179-84.
- 10 Czyrski A, Szalek E. An HPLC method for levofloxacin determination and its application in biomedical analysis. *J Anal Chem.* 2016;71(8):874-7.
- 11 Gupta H, Aqil M, Khar RK, et al. Development and validation of a stability-Indicating RP-UPLC method for the quantitative analysis of sparfloxacin. *J Chromatogr Sci.* 2010;48:1-6.
- 12 Razzaq SN, Ashfaq M, Khan IU, et al. Simultaneous determination of dexamethasone and moxifloxacin in pharmaceutical formulations using stability indicating HPLC method. *Arab J Chem.* 2017;10(3):321-8.